DIRECTORY OF SERVICES - ONCOGENETICS

FY- 2020 - 2021

Lilac Insights™

passion • commitment • genetics

Lilac Insights™

passion • commitment • genetics

INDEX

Hematopoietic and Lymphoid Malignancies

Solid Tumors

Hematopoietic and Lymphoid Malignancies

Lilac Insights™
passion • commitment • genetics

Acute Lymphoblastic Leukemia (ALL) Test panels

ACUTE LYMPHOBLASTIC LEUKEMIA (ALL) TEST PANELS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G10S07T01	**Oncoinsights™ Cytogenetic Panel B- ALL (Ploidy Analysis + FISH) + Molecular: *IKZF1* mutation analysis + FISH for ph1 like ALL**	Ploidy Analysis + FISH + Molecular	BMA of 3-5ml in Sodium Heparin + BMA 3-5ml in EDTA	**Ploidy Analysis** + **FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3,* [t(1;19)], *KMT2A (MLL)* translocation[t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17 And **FISH:** Ph1-Like ALL: *ABL1, ABL2, PDGFRB, JAK2 & CRLF2* translocations, **Molecular:** *IKZF1* mutation Analysis .	8 - 10
2	G10S07T02	**Cytogenetic Panel B- ALL (CK + FISH) + Molecular: *IKZF1* mutation analysis + FISH for ph1 like ALL**	Conventional Karyotyping + FISH + Molecular	BMA of 3-5ml in Sodium Heparin + BMA 3-5ml in EDTA	**Conventional Karyotyping, FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3,* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17 And **FISH:** Ph1-Like ALL: *ABL1, ABL2, PDGFRB, JAK2 & CRLF2* translocations, **Molecular:** *IKZF1* mutation Analysis.	8 - 10
3	G10S07T03	**Oncoinsights™ Cytogenetic panel. Ploidy Analysis + FISH B-ALL, Molecular: *IKZF1* mutation analysis Reflex to FISH for Ph1 Like ALL**	Ploidy Analysis + FISH + Molecular	BMA of 3-5ml in Sodium Heparin + BMA 3-5ml in EDTA	**Ploidy Analysis, +** **FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3,* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17, **Molecular** *IKZF1* mutation Analysis & in Reflex **FISH:** Ph1 like ALL: *ABL1, ABL2, PDGFRB, JAK2 & CRFL2* translocations.	8 - 10
4	G10S07T04	**Oncoinsights™ CK + FISH B-ALL, Molecular: *IKZF1* mutation analysis Reflex to FISH for Ph1 Like ALL**	Conventional Karyotyping + FISH + Molecular	BMA of 3-5ml in Sodium Heparin	**Conventional Karyotyping, + FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17, **Molecular** *IKZF1* mutation Analysis & in Reflex **FISH:** Ph1 like ALL: *ABL1, ABL2, PDGFRB, JAK2 & CRFL2* translocations	8 - 10
5	G09S22T02	**Oncoinsights™ Ph1 like ALL FISH panel extended**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** *ABL1, ABL2, PDGFRB, JAK2 & CRFL2* translocations	5
6	G10S07T06	**Oncoinsights™ Ph1 Like B-ALL FISH Panel Extended + *IKZF1* mutation analysis**	FISH+ Molecular	BMA of 3-5ml in Sodium Heparin	**FISH:** *ABL1, ABL2, PDGFRB, JAK2 & CRFL2* translocations, **Molecular** *IKZF1* mutation analysis	5
7	G09S07T09	**Oncoinsights™ FISH panel B - ALL**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* tarnslocation, +4, +10, +17	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Lymphoblastic Leukemia (ALL) Test panels

ACUTE LYMPHOBLASTIC LEUKEMIA (ALL) TEST PANELS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
8	G09S07T10	**Oncoinsights™ Cytogenetic panel B-ALL**	Ploidy Analysis + FISH	BMA of 3-5ml in Sodium Heparin	**Ploidy Analysis, +** **FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17	8 - 10
9	G09S07T14	**Oncoinsights™ CK + FISH B-ALL**	Conventional Karyotyping+ FISH	BMA of 3-5ml in Sodium Heparin	**Conventional Karyotyping,** **FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17	8 - 10
10	G09S07T15	**Conventional Karyotyping for B-ALL**	Conventional Karyotyping	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping for B-ALL**	10
11	G10S07T05	**Oncoinsights™ FISH panel B - ALL + *IKZF1* mutation analysis**	FISH + Molecular	BMA of 3-5ml in Sodium Heparin	**FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* tarnslocation, +4, +10, +17, **Molecular** *IKZF1* mutation analysis	5
12	G09S07T12	**Oncoinsights™ Cytogenetic panel B-ALL Reflex to Ph1 Like ALL**	Ploidy Analysis + FISH	BMA of 3-5ml in Sodium Heparin	**Ploidy Analysis, +** **FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* translocation, +4, +10, +17 in Reflex **FISH:** *ABL1, ABL2, PDGFRB, JAK2 & CRFL2* translocations	8-10
13	G09S07T13	**Oncoinsights™ CK + B-ALL Reflex to Ph1 Like ALL**	CK + FISH	BMA of 3-5ml in Sodium Heparin	**Conventional Karyotyping, +** **FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], +4, +10, +17 in Reflex **FISH:** *ABL1, ABL2, PDGFRB, JAK2 & CRFL2* translocations	8-10
14	G09S07T09	**Oncoinsights™ FISH panel B - ALL**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** *BCR-ABL1, ETV6-RUNX1, PBX1-TCF3* [t(1;19)], *KMT2A(MLL)* translocation [t(9;11), t(11;19), t(4;11)], *IGH* tarnslocation, +4, +10, +17	5
15	G08S07T08	**ALL Panel(deletion/duplications) by MLPA**	Molecular	BMA of 3-5ml in EDTA tube	**Molecular:** ALL Panel by *MLPA, loss of 5q, 6q, 7q, & IKZF1, MYC, MYCN, ALK, JAK2 (V617F),* deletions of *9p21.3, 13q, 17p, 17q* & *PAX5* gene, *PTEN, ATM, ETV6, RUNX1,* & chromosome 12, 18 &19.	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Lymphoblastic Leukemia (ALL) Test panels

ACUTE LYMPHOBLASTIC LEUKEMIA (ALL) TEST PANELS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
16	G08S07T06	**ALL Multiplex by PCR**	Molecular	BMA of 3-5ml in EDTA tube	**Molecular:** ALL Multiplex by *PCR*, t(1;19), t(9;22), t(12;21), t(4;11)	10
17	G08S07T05	***BCR-ABL1* by qualitative PCR**	Molecular	BMA/PB of 3-5ml in EDTA tube	**Molecular:** *BCR-ABL1* for p210 & p190 by Qualitative PCR	4
18	G08S07T02	***BCR-ABL1* by quantitative PCR (p210 & p190) & *ABL1* kinase domain mutation**	Molecular	BMA/PB of 3-5ml in EDTA tube	**Molecular:** *BCR-ABL1* RT-qPCR for p210 & p19, *ABL1* kinase domain mutation (sequencing)	7
19	G08S06T03	***BCR-ABL1* by quantitative PCR (p210 & p190)**	Molecular	BMA/PB of 3-5ml in EDTA tube	**Molecular:** *BCR-ABL1* RT-qPCR for p210 & p190	7
20	G09S21T01	**Oncoinsights™ FISH panel T - ALL**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** del(9p21), *KMT2A(MLL)* translocation, *TCR-A/D*(14q11) translocation, *TCR-B*(7q34) translocation, *TLX1*(10q24) translocation, *TLX3*(5q35) translocation/rearrangement.	5
21	G09S19T01	**Oncoinsights™ FISH panel T-Cell Prolymphocytic Leukemia (T-PLL)**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** *TCR-A/D* (14q11) translocation, i(8q), +8	5
22	G08S16T01	**Molecular Panel for ETP-ALL *(FLT3-ITD* and *D835)***	Molecular	BMA/PB of 3-5ml in EDTA tube	**Molecular:** *FLT3-ITD* and *D835*	3
23	G08S15T16	**TPMT Genotyping**	Molecular	BMA of 3-5ml in EDTA	**Molecular:** Thiopurine Methyltransferase Genotyping test	10

ACUTE LYMPHOBLASTIC LEUKEMIA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
24	G09S07T01	**Ploidy Analysis**	Cytogenetics Study	BMA of 3-5ml in Sodium Heparin	**Ploidy Analysis**	5
25	G09S10T06	**t(9;22)*(BCR-ABL1)***	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(9;22)*(BCR-ABL1)*	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Lymphoblastic Leukemia (ALL) Test panels

ACUTE LYMPHOBLASTIC LEUKEMIA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
26	G09S07T02	***NUP214*(9q34.13) translocation/ rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *NUP214*(9q34.13) translocation/ rearrangement	5
27	G09S07T03	***JAK2*(9p24) translocation/rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *JAK2*(9p24) translocation/rearrangement	5
28	G09S07T04	***TCF3* variant(v;19)**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *TCF3* variant*(v;19)*	5
29	G09S07T06	**Trisomy for 4,10 & 17 analysis**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** Trisomy for 4,10 & 17 analysis	5
30	G09S07T07	**t(1;19)** *PBX1-TCF3*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(1;19) *PBX1-TCF3*	5
31	G09S07T08	**t(12;21)** *ETV6-RUNX1*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(12;21) *ETV6-RUNX1*	5
32	G09S32T01	*TCR-A/D* **t(14q11) translocation**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *TCR-A/D* t(14q11) translocation	5
33	G09S32T02	*TCR-B* **t(7q34) translocation**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *TCR-B* t(7q34) translocation	5
34	G09S32T03	*TLX1:* **t(10q24) translocation**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *TLX1* t(10q24) translocation	5
35	G09S32T04	*TLX3:* **t(5q35) translocation**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *TLX3* t(5q35) translocation	5
36	G09S32T05	**del(9p21): p16/***CDKN2A* **deletion**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** del(9p21): p16/*CDKN2A* deletion	5
37	G09S35T01	*KMT2A (MLL)* **translocation**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *KMT2A(MLL)* translocation	5
38	G09S34T01	**t(9;11)** *(MLLT3-KMT2A)*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(9;11)*(MLLT3-KMT2A)*	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Lymphoblastic Leukemia (ALL) Test panels

ACUTE LYMPHOBLASTIC LEUKEMIA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
39	G09S34T02	**t(4;11)***(AFF1-KMT2A)*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(4;11) *(AFF1-KMT2A)*	5
40	G09S34T03	**t(11;19)***(MLLT1-KMT2A)*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(11;19) *(MLLT1-KMT2A)*	5
41	G09S34T04	**t(6;11)***(MLLT4-KMT2A)*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** t(6;11) *(MLLT4-KMT2A)*	5
42	G09S07T05	*ABL1* **(9q34) translocation/rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *ABL1* (9q34) translocation/rearrangement	5
43	G09S22T03	*ABL2* **translocation/rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *ABL2* translocation/rearrangement	5
44	G09S22T04	*PDGFRB* **translocation/rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *PDGFRB* translocation/rearrangement	5
45	G09S22T05	*CRLF2* **translocation/rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** *CRLF2* translocation/rearrangement	5
46	G09S07T16	**iAMP21**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH :** iAMP21	10
47	G08S07T01	*ABL1* **Kinase domain mutation**	Molecular	BMA of 3-5ml in EDTA	**Molecular:** *ABL1* Kinase domain mutation	7
48	G08S07T03	*BCR-ABL1* **RT-qPCR for p210 (IS)**	Molecular	BMA of 6-10ml in EDTA	**Molecular:** *BCR-ABL1* RT-qPCR for p210 (IS)	4
49	G08S07T04	*BCR-ABL1* **RT-qPCR for p190**	Molecular	BMA of 6-10ml in EDTA	**Molecular:** *BCR-ABL1* RT-qPCR for p190	4
50	G08S07T05	*BCR-ABL1* **breakpoint analysis by Qualitative PCR**	Molecular	BMA of 3-5ml in EDTA	**Molecular:** *BCR-ABL1* breakpoint analysis by Qualitative PCR	4
50	G08S07T07	*IKZF1* **alteration(exon 2-8) deletion**	Molecular	BMA of 3-5ml in EDTA	**Molecular:** *IKZF1* Alteration	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Myeloid Leukemia Test panels

ACUTE MYELOID LEUKEMIA TEST PANELS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G10S01T02	**Oncoinsights™ Advanced Combipanel Panel - AML**	Conventional Karyotyping + FISH + NGS	BMA of 3ml in Sodium Heparin Tubes(2) + BMA of 3ml in EDTA tube(1)	**Conventional Karyotyping +** **FISH:** t(8;21), *PML-RARA*, inv(16), *BCR-ABL1*, *KMT2A(MLL)* translocation, -5/del(5q), -7/del(7q), +8, *3q26 (MECOM)* translocation, del(17p) + **NGS:** HOTSPOT GENES: 23 genes, FULL GENES: 17 genes	10-12
2	G10S01T01	**Oncoinsights™ combipnel - AML**	Conventional Karyotyping + FISH + Molecular	BMA of 3ml in Sodium Heparin Tubes(2) + BMA of 3ml in EDTA tube(1)	**Conventional Karyotyping +** **FISH:** t(8;21), *PML-RARA*, inv(16), *BCR-ABL1*, *KMT2A(MLL)* translocation, -5/del(5q), -7/del(7q), +8, 3q26*(MECOM)* translocation, del(17p) + **Molecular** - *FLT3 - ITD* (Fragment Analysis), *FLT3-TKD* (sequencing), *CEBPA, NPM1* mutation by sequencing	8-10
3	G10S01T04	**Oncoinsights™ Conventional Karyotyping + NGS AML**	Conventional Karyotypig + NGS	BMA of 3ml in Sodium Heparin Tubes(2) + BMA of 3ml in EDTA tube(1)	**Conventional Karyotyping +** **NGS:** HOTSPOT GENES: 23 genes, Full Genes: 17 genes .	8-12
4	G10S01T03	**Oncoinsights™ NGS Panel AML (Mutations & Translocations)**	NGS	BMA/PB of 3-5ml in EDTA	**NGS:** HOTSPOT GENES: 23 genes, Full genes: 17 RNA - Fusion Driver genes: *ABL1, ALK, BCL2, BRAF, CCND1, CREBBP, EGFR, ETV6(TEL), FGFR1, FGFR2, FUS, HMGA2, JAK2, KMT2A(MLL), MECOM, MET, MLLT10, MLLT3, MYBL, MYH11, NTRK3, NUP214, PDGFRA, PDGFRB, RARA, RBM15, RUNX1(AML1), TCF3(E2A) TFE3.*	12-15
5	G09S09T08	**Oncoinsights™ Cytogenetic Panel AML**	Conventional Karyotyping + FISH	BMA of 3ml in Sodium Heparin Tubes(2)	**Conventional Karyotyping +** **FISH:** t(8;21), *PML-RARA*, inv(16), *BCR-ABL1*, *KMT2A(MLL)* translocation, -5/del(5q), -7/del(7q), +8, *3q26 (MECOM)* translocation, del(17p)	8-10
6	G09S09T09	**Oncoinsights™ FISH Panel - AML**	FISH	BMA of 3ml in Sodium Heparin	**FISH:** t(8;21), *PML-RARA*, inv(16), *BCR-ABL1*, *KMT2A(MLL)* translocation, -5/del(5q), -7/del(7q), +8, *3q26 (MECOM*) translocation, del(17p)	5
7	G08S01T04	**Molecular Panel AML**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular** *FLT3(ITD)* by Fragment Analysis, *FLT3-TKD, CEBPA, NPM1* by Sequencing	3

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Myeloid Leukemia Test panels

ACUTE MYELOID LEUKEMIA TEST PANELS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
8	G08S01T06	***KIT* Molecular Panel AML**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *KIT*(exon17)*(PCR)* and *KIT*(exon8) by Sequencing	5
9	G08S01T07	***FLT3* Molecular Panel AML**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *FLT3(ITD)* by Fragment Analysis, *FLT3-TKD* by Sequencing	9
10	G08S01T11	***IDH1* & *IDH2* by Sequencing**	Molecular	BMA of 3-5ml in EDTA	**Molecular:** *IDH1* & *IDH2* by Sequencing	6
11	G08S01T01	**Oncoinsights™ Extended Molecular Panel**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *FLT3-ITD & TKD, NPM1, CEBPA, ASXL1, DNMT3A, IDH1 & IDH2, TP53* mutation by sequencing	7
12	G08S01T14	**MRD for t(8;21)(Real time PCR)**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** MRD for t(8;21)(Real time PCR)	10
13	G08S15T03	**Oncoinsights™ NGS Panel AML(Mutations)**	NGS	BMA/PB of 3-5ml in EDTA	**NGS:** Myeloid NGS - DNA Studies: HOTSPOT GENES :23 genes. FULL GENES 17 genes	10-12
14	G08S15T15	**Oncoinsights™ NGS Panel AML(FUSIONS)**	NGS	BMA/PB of 3-5ml in EDTA	**NGS:** Myeloid NGS - RNA Studies: Fusion Driver genes: *ABL1, ALK, BCL2, BRAF, CCND1, CREBBP, EGFR, ETV6(TEL), FGFR1, FGFR2, FUS, HMGA2, JAK2, KMT2A(MLL), MECOM, MET, MLLT10, MLLT3, MYBL, MYH11, NTRK3, NUP214, PDGFRA, PDGFRB, RARA, RBM15, RUNX1(AML1), TCF3(E2A) TFE3.*	10-12

ACUTE MYELOID LEUKEMIA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
15	G09S33T01	**t(9;22):** *BCR-ABL1*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** t(9;22): *BCR-ABL1*	5
16	G09S33T02	*KMT2A(MLL)* **translocation**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** *KMT2A(MLL)* translocation	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Myeloid Leukemia Test panels

ACUTE MYELOID LEUKEMIA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
17	G09S33T03	**t(9;11)***(MLLT3-KMT2A)*, **t(4;11)***(AFF1-KMT2A)*, **t(11;19)***(MLLT1-KMT2A)*, **t(6;11)** *(MLLT4-KMT2A)*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** t(9;11)*(MLLT3-KMT2A)*, t(4;11)*(AFF1-KMT2A)*, t(11;19)*(MLLT1-KMT2A)*, t(6;11)*(MLLT4-KMT2A)*	5
18	G09S33T04	**-5/del(5q)**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** -5/del(5q)	5
19	G09S33T05	**-7/del(7q)**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** -7/del(7q)	5
20	G09S33T07	**-20/del(20q)**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** -20/del(20q)	5
21	G09S33T06	**trisomy** *8*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** trisomy 8	5
22	G09S33T08	**trisomy 21**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** trisomy 21	5
23	G09S09T01	**Conventional Karyotyping for AML**	Conventional Karyotyping	BMA of 3-5ml in Sodium Heparin	**Conventional Karyotyping for AML**	10-12
24	G09S09T02	**t(1;22):** *RBM15-MKL1*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** t(1;22): *RBM15-MKL1*	5
25	G09S09T03	**t(6;9):** *DEK-NUP214*	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** t(6;9): *DEK-NUP214*	5
26	G09S09T04	**inv(3):** *MECOM* **(EVI) translocation/ rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** inv(3): *MECOM* translocation/ rearrangement	5
27	G09S09T05	**trisomy 8**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** trisomy 8	5
28	G09S09T06	**inv(16):** *CBF-B* **rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** inv(16): *CBF-B* rearrangement	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute Myeloid Leukemia Test panels

ACUTE MYELOID LEUKEMIA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
29	G09S09T07	**t(8;21):** *RUNX1-RUNX1T1* **rearrangement**	FISH	BMA of 3-5ml in Sodium Heparin	**FISH:** t(8;21) : *RUNX1-RUNX1T1*	5
30	G08S01T02	*c-KIT D816V* **mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *c-KIT (D816V)* mutation	5
31	G08S01T03	*FLT3-ITD* **Mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *FLT3-ITD* Mutation	3
32	G08S01T05	*NPM1* **mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *NPM1* mutation	3
33	G08S01T08	*FLT3-D835* **mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *FLT3-D835(TKD)* mutation	5
34	G08S01T09	*CEBPA* **mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *CEBPA* mutation	3
35	G08S01T10	*c-KIT* **Exon 8 mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *c-KIT* Exon 8 mutation	5
36	G08S01T12	*DNMT3A(R882)* **mutation**	Molecular	BMA/PB of 3-5ml in EDTA	**Molecular:** *DNMT3A(R882)* mutation	4
37	G08S02T04	**RT-qPCR for** *PML-RARA* **transcript quantitaive analysis (***BCR3***)**	Molecular	BMA of 3-5ml in EDTA Tube	**Molecular:** *PML-RARA* transcript copies(quantitative) RT-qPCR(*BCR1*)	5
38	G08S02T02	**RT-qPCR for** *PML-RARA* **transcript quantitaive analysis (***BCR3***)**	Molecular	BMA of 3-5ml in EDTA Tube	**Molecular:** *PML-RARA* transcript copies(quantitative) RT-qPCR*(BCR3)*	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Acute promyelocytic Leukemia (APL)Test panels

ACUTE PROMYELOCYTIC LEUKEMIA (APL)TEST PANELS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S25T01	**FISH for APL or FISH for APML**	FISH	BMA of 3-5ml in Sodium Heparin Tube	**FISH:** *PML-RARA* t(15;17)	5
2	G09S25T02	***RARA* Variant Analysis**	FISH	BMA of 3-5ml in Sodium Heparin Tube	**FISH:** t(v;17)	5
3	G08S02T01	**Qualitative Analysis for *PML-RARA***	Molecular	BMA of 3-5ml in EDTA Tube	**Molecular:** *PML-RARA* transcript analysis (qualitative)	4
4	G08S02T05	**RT-qPCR for *PML-RARA* transcript quantitaive analysis *(BCR1)***	Molecular	BMA of 3-5ml in EDTA Tube	**Molecular:** *PML-RARA* transcript copies (quantitative) RT-qPCR *(BCR1 & BCR3)*	5

Bone Marrow Transplant Test Menu

BONE MARROW TRANSPLANT TEST MENU						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S05T01	**Sex-mismatched Post Bone marrow transplant monitoring**	CK + FISH	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping** + **FISH:** XX/XY	5
2	G09S25T02	**Sex-mismatched Post Bone Marrow transplant monitoring**	FISH	BMA of 3-5ml in Sodium Heparin Tube	**FISH:** XX/XY	5
3	G09S05T03	**Post- BMT Case Conventional Karyotyping(Bone Marrow Transplant)**	Conventional Karyotyping	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping**	10-12
4	G08S03T03	**STR Chimerism Analysis**	PCR	PB of 3ml in EDTA	**Molecular:** STR Chimerism Analysis	10-12
5	G08S03T01	**HLA Typing High Resolution Test**	NGS	PB of 5ml in EDTA 2 tubes	**Molecular:** HLA Typing High Resolution Test	12

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Chronic Eosinophilic Leukemia

CHRONIC EOSINOPHILIC LEUKEMIA (CEL)						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S23T01	**Oncoinsights™ FISH Panel CEL**	FISH	BMA 3-5 ml in Sodium Heparin tube	**FISH:** *PDGFRA* (4q12) translocation with *CHIC2* deletion, *PDGFRB* (5q33) translocation, *FGFR1* (8p12) translocation	5

B-Cell & T-Cell Lymphoma

B-CELL AND T-CELL LYMPHOMA						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S13T03	**FISH for *IGH* Translocation (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** *IGH* Translocation	7
2	G09S13T02	**FISH for *IGH* Translocation (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** *IGH* Translocation	5
3	G09S13T05	**FISH for Mantle Cell Lymphoma (*MCL*) (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** t(11;14)(*IGH/CCND1*)	7
4	G09S13T04	**FISH for Mantle Cell Lymphoma (*MCL*) (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(11;14)(*IGH/CCND1*)	5
5	G09S13T07	**FISH for Follicular Lymphoma (FL)(*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** t(14;18)*(IGH/BCL2)*	7
6	G09S13T06	**FISH for Follicular Lymphoma(FL)(*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(14;18)*(IGH/BCL2)*	5
7	G09S13T09	**FISH for Burkitt Lymphoma(*FFPE* Block)**	FISH	FFPE Tissue Blockion)	**FISH:** t(8q24)(*MYC* translocation)	7
8	G09S13T08	**FISH for Burkitt Lymphoma (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(8q24)(*MYC* translocation)	5
9	G09S13T11	**FISH for Diffused Large B-cell Lymphoma (*DLBCL*)(*FFPE* block)**	FISH	FFPE Tissue Block	**FISH:** t(14;18)(*IGH/BCL2*), t(3q27)(*BCL6* translocation)	7
10	G09S13T10	**FISH for Diffused Large B-cell Lymphoma (*DLBCL*)(*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(14;18)(IGH/BCL2), t(3q27)(*BCL6* translocation)	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

B-Cell & T-Cell Lymphoma

B-CELL LYMPHOMA AND T-CELL LYMPHOMA						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
11	G09S13T13	**FISH for Dual Hit Lymphoma (High Grade B -cell Lymphoma)(*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** t(14;18)(*IGH/BCL2*), t(8q24)(*MYC* translocation)	7
12	G09S13T12	**FISH for Dual Hit Lymphoma (High Grade B -cell Lymphoma)(*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(14;18)*(IGH/BCL2), MYC(8q24)* translocation	5
13	G09S13T15	**Oncoinsights™FISH Panel for Triple Hit Lymphoma(High Grade B-cell Lymphoma) (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** t(14;18)*(IGH/BCL2), MYC(8q24)* translocation, t(3q27) (*BCL6* translocation)	7
14	G09S13T14	**Oncoinsights™FISH Panel for Triple Hit Lymphoma(High Grade B-cell Lymphoma) (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(14;18)*(IGH/BCL2), MYC(8q24)* translocation, t(3q27)(*BCL6* translocation)	5
15	G09S13T17	**FISH for MALT Lymphoma(*FFPE* Block)**	FISH	FFPE Block	**FISH:** t(11;18)(*MALT1*) translocation, t(14;18) (*IGH/MALT1*), t(3q27)(*BCL6* translocation)	7
16	G09S13T16	**FISH for MALT Lymphoma (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(11;18)(*MALT1*) translocation, t(14;18) (*IGH/MALT1*), t(3q27)(*BCL6* translocation)	5
17	G09S13T19	**FISH for Lymphoma B-Cell (*FFPE* Block)**	FISH	FFPE Block	**FISH:** t(11;14)(*IGH/CCND1*), t(14;18)(*IGH/BCL2*), t(8q24)(*MYC* translocation), t(3q27)(*BCL6* translocation)	7
18	G09S13T18	**FISH for Lymphoma B-Cell (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(11;14)(*IGH/CCND1*), t(14;18)(*IGH/BCL2*), t(8q24)(*MYC* translocation), t(3q27)(*BCL6* translocation)	5
19	G09S13T21	**FISH for Lymphoma T-Cell (*FFPE* Block)**	FISH	FFPE Block	**FISH:** t(2p23)(*ALK* translocation), i(7q), +8	7
20	G09S13T20	**FISH for Lymphoma T-Cell (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** t(2p23)(*ALK* translocation), i(7q), +8	5
21	G09S13T23	**Oncoinsights™ FISH Panel Hepatosplenic γ, δ T-cell Lymphoma (*FFPE* Block)**	FISH	FFPE Block	**FISH:** : i(7q), +8	7
22	G09S13T22	**Oncoinsights™ FISH Panel Hepatosplenic γ, δ T-cell Lymphoma (*BMA/PB*)**	FISH	BMA/PB of 3-5ml in Sodium Heparin tube	**FISH:** : i(7q), +8	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

B-Cell & T-Cell Lymphoma

B-CELL LYMPHOMA AND T-CELL LYMPHOMA						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
23	G09S13T01	**Conventional Karyotyping (Lymphoma)**	Karyotyping	BMA/PB of 3-5ml in Sodium Heparin tube	**Conventional Karyotyping – Lymphoma**	8-10
24	G08S15T23	***TP53* Sequencing on *FFPE***	Molecular	FFPE Block	**Molecular:** *TP53* (exons 4-9) by sanger Sequencing	10
25	G08S15T01	***TP53* Sequencing on *BMA/PB***	Molecular	3-5ml of BMA/PB in EDTA tube	**Molecular:** *TP53* (exons 4-9) by sanger Sequencing	10

B-CELL LYMPHOMA AND T-CELL LYMPHOMA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
26	G09S13T25	**FISH for *8q24(MYC)* Translocation (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** 8q24*(MYC)* Translocation (*FFPE* Block)	7
27	G09S13T24	**FISH for *8q24(MYC)* Translocation(BM)**	FISH	BMA of 3-5ml in Sodium Heparin tube	**FISH:** 8q24*(MYC)* Translocation(BM)	5
28	G09S13T27	**FISH for Lymphoma *2p23 (ALK)* translocation (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** Lymphoma 2p23*(ALK)* translocation (*FFPE* Block)	7
29	G09S13T26	**FISH for Lymphoma *2p23 (ALK)* translocation (BM)**	FISH	BMA of 3-5ml in Sodium Heparin tube	**FISH:** Lymphoma 2p23*(ALK)* translocation (BM)	5
30	G09S13T29	**FISH for Lymphoma t(3q27)(*BCL6*) translocation (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** Lymphoma t(3q27)(*BCL6*) translocation (*FFPE* Block)	7
31	G09S13T28	**FISH for Lymphoma t(3q27)(*BCL6*) translocation (BM)**	FISH	BMA of 3-5ml in Sodium Heparin tube	**FISH:** Lymphoma t(3q27)(*BCL6*) translocation (BM)	5
32	G09S13T31	**FISH for Lymphoma t(11;18)(*MALT1*) translocation (*FFPE* Block)**	FISH	FFPE Tissue Block	**FISH:** Lymphoma t(11;18)(*MALT1*) translocation (*FFPE* Block)	7
33	G09S13T30	**FISH for Lymphoma t(11;18)(*MALT1*) translocation (BM)**	FISH	BMA of 3-5ml in Sodium Heparin tube	**FISH:** Lymphoma t(11;18)(*MALT1*) translocation (BM)	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Chronic Lymphocytic Leukemia

CHRONIC LYMPHOCYTIC LEUKEMIA (CLL) PANEL						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G10S02T01	**Oncoinsights™ Combipanel CLL**	CK + FISH + Molecular	BMA/PB 3-5 ml in Sodium Heparin tube and EDTA tube	**Conventional Karyotyping** by *B-Cell* Mitogen stimulation, **FISH:** del(13q), del(11q) , del(6q), del(17p), +12, t(11;14), **Molecular** *IGHV* mutations, *TP53* (exons 4-9)(sequencing)	8-10
2	G09S06T04	**Oncoinsights™ Cytogenetic Panel CLL**	CK + FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping** by *B-Cell* Mitogen Stimulation, **FISH:** del(13q), del(11q), del(6q), del(17p), +12, t(11;14)	8-10
3	G09S06T05	**Oncoinsights™ FISH Panel CLL**	FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**FISH:** del(13q), del(11q), del(6q), del(17p), +12, t(11;14)	5
4	G08S05T02	**Oncoinsights™ Molecular Panel CLL**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *NOTCH1* mutation(p2514Fs*4)(PCR), *TP53* (exons 4-9) by sequencing	10
5	G08S05T04	**Oncoinsights™ Molecular Panel CLL (*IGHV&TP53*)**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *IGHV* mutations, *TP53* (exons 4-9) by sequencing	10

CHRONIC LYMPHOCYTIC LEUKEMIA (CLL) INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
6	G09S31T01	**-13/del(13q) Analysis**	FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**FISH:** -13/del(13q) Analysis	5
7	G09S31T02	**del(17p) Analysis**	FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**FISH:** del(17p) Analysis	5
8	G09S31T03	**del(11q) Analysis**	FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**FISH:** del(11q) Analysis	5
9	G09S06T03	**Trisomy 12 Analysis**	FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**FISH:** Trisomy 12 Analysis	5
10	G09S06T02	**14q32*(IGH)* Translocation**	FISH	BMA/PB 3-5 ml in Sodium Heparin tube	**FISH:** 14q32*(IGH)* Translocation	5
11	G09S06T01	**Conventional Karyotyping for CLL (*B-Cell* Stimulation)**	Conventional Karytoping	BMA/PB 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping** for *CLL* (*B-Cell* Stimulation)	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Chronic Lymphocytic Leukemia

CHRONIC LYMPHOCYTIC LEUKEMIA (CLL) INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
12	G08S05T01	*NOTCH1* **mutation (p2514Fs*4)(PCR)**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *NOTCH1* mutation(p2514Fs*4)(PCR)	5
13	G08S05T03	*IGHV* **mutations**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *IGHV* mutations	10
14	G08S05T05	*TP53* **(exons 4-9) mutation**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *TP53* (exons 4-9) by sequencing	10

Chronic Myeloid Leukemia

CHRONIC MYELOID LEUKEMIA(CML)						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G10S03T01	**Oncoinsights™ Combipanel chronic phase (CP)**	CK + FISH + Molecular	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping** + **FISH:** *BCR-ABL1* + **Molecular:** *BCR-ABL1* RT-qPCR for *p210(IS)*	7
2	G09S10T02	**Oncoinsights™ Cytogenetic Panel CML - Chronic Phase (CP)**	CK + FISH	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping** + **FISH:** *BCR-ABL1*	7
3	G09S10T01	**Oncoinsights™ Cytogenetic Panel CML - Blastic Phase (BP)**	CK + FISH	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping** + **FISH:** *BCR-ABL1*, +8, +19, +21, del(17p)	8-10
4	G09S10T03	**Conventional Karyotyping**	Karyotyping	BMA/PB 3-5 ml in sodium heparin tube	**Conventional Karyotyping**	5-7
5	G09S10T04	**FISH for CML- Chronic Phase (CP)**	FISH	BMA 3-5 ml in Sodium Heparin tube	**FISH:** *BCR-ABL1*	5
6	G09S10T05	**Oncoinsights™ FISHPanel CML- Blastic Phase (BP)**	FISH	BMA 3-5 ml in Sodium Heparin tube	**FISH:** *BCR-ABL1*, +8, +19, +21, del(17p)	5
7	G09S10T07	**Oncoinsights™ FISH for** *BCR-ABL1* **(***FFPE* **Block)**	FISH	FFPE Block	**FISH:** *BCR-ABL1*	7
8	G08S06T06	**Molecular Panel for CML**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *BCR-ABL1* RT-qPCR for *p210(IS)* &/or p190, *ABL1* Kinase domain mutation	7

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Chronic Myeloid Leukemia

CHRONIC MYELOID LEUKEMIA(CML)						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
9	G08S06T02	**RT-qPCR for *BCR-ABL1* transcript copies *p210(IS)*, Quantitative Analysis**	Molecular	BMA/PB 5-7 ml in EDTA tube	**Molecular:** *BCR-ABL1* RT-qPCR for *p210(IS)*	4
10	G08S06T04	***ABL1* Kinase domain mutation**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *ABL1* Kinase domain mutation	7
11	G08S06T03	***BCR-ABL1* RT-qPCR for *p210(IS)* &/or *p190***	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *BCR-ABL1* RT-qPCR for *p210(IS)* &/or *p190*	7
12	G08S06T07	***BCR-ABL1* RT-qPCR for *p190***	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *BCR-ABL1* RT-qPCR for *p190*	7
13	G08S06T01	***BCR-ABL1* Qualitative Analysis**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *BCR-ABL1* transcript by breakpoint analysis (qualitative PCR)	4
14	G09S10T06	**FISH for t(9;22)**	FISH	BMA 3-5ml in Sodium Heparin	**FISH:** t(9;22)	5

Conventional Karyotyping, Constitutional Karyotyping, Cell line Karyotyping, Chromosomal Breakage studies

CONVENTIONAL KARYOTYPING, CONSTITUTIONAL KARYOTYPING, CELL LINE KARYOTYPING, CHROMOSOMAL BREAKAGE STUDIES						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S11T05	**Conventional Karyotyping**	Conventional Karyotyping	BMA 3-5 ml in Sodium Heparin tube	**Conventional Karyotyping**	10
2	G09S11T04	**Constitutional Karyotyping**	Conventional Karyotyping	BMA 3-5 ml in Sodium Heparin tube	**Constitutional Karyotyping**	10
3	G09S11T03	**OncoInsights™ Cell Line Karyotyping**	Conventional Karyotyping	Suspension/ Monolayer Culture	**Cell line Karyotyping**	15
4	G09S11T06	**Oncoinsights™ Chromosomal Breakage studies in Fanconi's Anemia/Aplastic Anemia**	Spontaneous and MMC, DEB-induced Chromosomal Breakage studies	PB 5-7 ml of patient and 5-7ml PB of unrelated age-sex matched control in Sodium Heparin tube	**Chromosomal Breakage studies**	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Hairy Cell Leukemia

Hairy cell leukemia (HCL)						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S18T01	**Oncoinsights™ FISH panel HCL**	FISH	BMA 3-5 ml in Sodium Heparin tube	**FISH:** *IGH, TCR-A/D*(14q11) translocation, -7/del(7q)	5
2	G08S08T01	**Molecular test HCL (*BRAFV600E* mutation)**	Molecular	BMA/PB 3-5 ml in EDTA tube	**Molecular:** *BRAF V600E* mutation *(ASO-PCR)*	6

Multiple Myeloma

Multiple Myeloma						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S20T09	**Oncoinsights™ Cytogenetic panel (MM) Multiple Myeloma**	FISH +CK	BMA 5-6ml in Sodium Heparin tube	**Conventional Karyotyping +** **FISH:** del(17p), -13/del (13q), del(1p), Amp(1q), *IGH* translocation , t(11;14), t(4;14), t(14;16), t(14;20), t(6;14), +3, +5,+7, +9, +15 (hyperdiploidy)	8-10
2	G09S20T01	**Oncoinsights™ FISH Panel MM Multiple Myeloma**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** del(17p), -13/del(13q), del(1p), Amp(1q), *IGH* translocation , t(11;14), t(4;14), t(14;16), t(14;20), t(6;14), +3, +5,+7, +9, +15 (hyperdiploidy)	5
3	G08S11T02	*MYD88 (L265P)*	Molecular	BMA of 3-5ml in EDTA tube	**Molecular:** *MYD288 (L265P)*	6

MULTIPLE MYELOMA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
4	G09S20T03	**del(17p) analysis**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** del(17p) analysis	5
5	G09S20T04	*IGH/CCND3:* **t(6;14) Analysis**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** *IGH/CCND3*: t(6;14) Analysis	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Multiple Myeloma

MULTIPLE MYELOMA INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
6	G09S20T05	*IGH/MAFB:* **t(14:20) Analysis**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** *IGH/MAFB*:t(14:20) Analysis	5
7	G09S20T06	*IGH/MAF:* **t(14:16) Analysis**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** *IGH/MAF*: t(14:16) Analysis	5
8	G09S20T07	*IGH/FGRF3:* **t(4;14) Analysis**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** *IGH/FGRF3*: t(4;14)	5
9	G09S20T08	**1q Amplification, 1p deletion analysis**	FISH	BMA 5-6ml in Sodium Heparin tube	**FISH:** 1q amplification, 1p deletion	5

Myelodysplastic Syndromes

MYELODYSPLASTIC SYNDROMES						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G10S04T03	**Oncoinsights™ Advanced Combipanel - MDS**	Conventional Karyotyping + FISH + NGS	BMA of 3-5 ml in Sodium Heparin and EDTA Tubes	**Conventional Karyotyping** + **FISH:** -5/del (5q),-7/del (7q), +8,del (20q), del (17p) + **NGS:** HOTSPOT GENES : 23 genes, FULL GENES:17 genes.	10-12
2	G10S04T04	**Oncoinsights™ MDS Conventional Karyotyping + NGS**	Conventional Karyotyping + NGS	BMA of 3-5 ml in Sodium Heparin and EDTA Tubes	**Conventional Karyotyping** + **NGS:** HOTSPOT GENES : 23 genes, FULL GENES:17 genes.	10-12
3	G08S15T03	**Oncoinsights™ MDS NGS (Mutations) Panel**	NGS	BMA of 3-5 ml in EDTA Tubes	**NGS:** hotspot genes - 23 genes, full genes: 17 genes.	10-12
4	G10S04T01	**Oncoinsights™ Combi-panel MDS**	Conventional Karyotyping+ FISH+Molecular	BMA 3-5ml in Sodium Heparin tube + 3-5ml in EDTA tube	**Conventional Karyotyping** + **FISH:** -5/del (5q), -7/del (7q), +8,del (20q), del (17p)+ **Molecular:** *ASXL1, DNMT3A, TP53* (exons 4-9) mutation	08-10
5	G09S08T01	**Oncoinsights™ Cytogenetic panel MDS**	Conventional Karyotyping+ FISH	BMA 3-5ml in Sodium Heparin tube	**Conventional Karyotyping** + **FISH:** -5/del (5q), -7/del (7q), +8,del (20q), del (17p)	08-10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Myelodysplastic Syndromes

MYELODYSPLASTIC SYNDROMES						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
6	G09S08T03	**Conventional Karyotyping for MDS**	Conventional Karyotyping	BMA 3-5ml in Sodium Heparin tube	**Conventional Karyotyping**	8-10
7	G09S08T03	**Oncoinsights™ FISH panel MDS**	FISH	BMA 3-5ml in Sodium Heparin tube	**FISH** -5/del(5q), -7/del (7q), +8, del (20q), del (17p)	5
8	G08S09T07	**Oncoinsights™ Molecular panel MDS**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *ASXL1*, *TP53* (exons 4-9), *DNMT3A* mutation	10

MYELODYSPLASTIC SYNDROMES INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
9	G08S09T02	*SF3B-1* **(exon14-16) mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *SF3B-1* (exon14-16) by sequencing	5
10	G08S09T03	*IDH1 & IDH2* **mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *IDH1 & IDH2* by Sequencing	6
11	G08S09T04	*MYD88 L265P* **mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *MYD88 L265P* mutation	6
12	G08S09T05	*ASXL1* **mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *ASXL1* by Sequencing	5
13	G08S09T06	*DNMT3A(R882)* **mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *DNMT3A(R882)* mutation	4
14	G08S09T08	*TP53* **mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *TP53* mutation(exon 4-9)	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Myeloproliferative Neoplasms, Myeloproloiferative Disorders

MYELOPROLIFERATIVENEOPLASMS/MYELOPROLIFERATIVE DISORDERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S14T05	**Oncoinsights™ FISH panel MPN/MPD**	FISH	BMA/PB 3-5ml in Sodium Heparin tube	**FISH** -7/*del(7q), +8, del (20q)*, *PDGFRA* (4q12) rearrangement with *CHIC2* deletion, *PDGFRB* (5q33) tarnslocation	5
2	G08S10T03	**Oncoinsights™** *JAK2* **Reflex Panel MPN/MPD**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *JAK2 V617F* mutation *(ASO-PCR)* and *JAK2* (exon12) by sequencing	5
3	G08S10T06	**Oncoinsights™ Molecular Panel** MPN/MPD *JAK2 V617F, MPL & CALR*	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *JAK2 V617F, MPL W515* mutation *(ASO-PCR), CALR* (exon 9)(sequencing)	5
4	G08S10T05	**Oncoinsights™ Molecular Panel MPN/MPD** *JAK2* **Exon 12**, *MPL & CALR*	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *JAK2* (exon 12), *MPL W515* mutation (ASO-PCR), *CALR* (exon 9)(sequencing)	5
5	G08S10T04	**Oncoinsights™ Molecular Panel MPN/MPD** *JAK2 reflex, MPL & CALR*	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *JAK2 V617F, JAK2* (exon 12), *MPL W515* mutation *(ASO-PCR)*, *CALR* (exon 9) (sequencing)	5
6	G08S10T09	**MPN Panel with** *BCR-ABL1:*	Molecular	BMA/PB 3-5ml in EDTA tube (Lavendar Top)	**Molecular:** *BCR-ABL1* by Qualitative PCR, *JAK2* (Exon-14(*V617F*) & Exon -12), *CALR, MPL*	5

MYELOPROLIFERATIVENEOPLASMS/MYELOPROLIFERATIVE DISORDERS INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
7	G09S14T01	**Myeloproliferative Neoplasms - Conventional Karyotyping**	Conventional Karyotyping	BMA/PB 3-5ml in Sodium Heparin tube	Myeloproliferative Neoplasms - **Conventional Karyotyping**	10
8	G09S14T02	*PDGFRB* **(5q33) rearrangement**	FISH	BMA/PB 3-5ml in Sodium Heparin tube	**FISH:** *PDGFRB* (5q33) translocation/rearrangement	5
9	G09S14T03	*PDGFRA* **(4q12) rearrangement with** *CHIC2* **deletion,**	FISH	BMA/PB 3-5ml in Sodium Heparin tube	**FISH:** *PDGFRA* (4q12) rearrangement with *CHIC2* deletion	5
10	G09S14T04	*FGFR1***(8p12) rearrangement**	FISH	BMA/PB 3-5ml in Sodium Heparin tube	**FISH:** *FGFR1*(8p12)translocation/ rearrangement	5
11	G08S10T01	*JAK2 V617F* **mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *JAK2 V617F* mutation	5
12	G08S10T08	*JAK2* **Exon-12 mutation**	Conventional Karyotyping	BMA/PB 3-5ml in Sodium Heparin tube	**Molecular:** *JAK2* Exon-12 mutation	5
13	G08S10T02	*CALR* **(Exon-9) mutation**	Molecular	BMA/PB 3-5ml in EDTA tube	**Molecular:** *CALR*(Exon-9) mutation by sequencing	5
14	G08S10T07	*MPL* **Mutation**	Molecular	BMA/PB 3-5ml in Sodium Heparin tube	**Molecular:** *MP*L Mutation	5

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Flow Cytometry

Lilac Insights™
passion • commitment • genetics

Flowcytometry

BONE MARROW ASPIRATE STUDIES						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G11S01T01	**Bone Marrow Advance**	Multiple	CBC sample, Aspirate Slides, Imprint Slides, Liquid BM Samples, Bone Marrow Biospy in formalin/AZF solution.	**Multiple: CBC Advance** (CBC, Peripheral Smear Review, Reticulocyte Count, Immature Platelet fraction, Reticulocyte Hemoglobin, Immature Reticulocyte Fraction), Bone Marrow Aspirate, Iron Staining, **Bone marrow Biopsy with Reflex IHC**	1-2 & 8 for BMB with reflex IHC

BONE MARROW ASPIRATE STUDIES						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G11S01T02	**Flowcytometry for leukemia's**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Leukemias	1-2
2	G11S01T03	**Flowcytometry for lymphoma's**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Lymphomas	1-2
3	G11S01T04	**Flowcytometry for myeloma**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Myeloma	1-2
4	G11S01T05	**Flowcytometry for Minimal Residual Disease (*MRD*) for B-ALL**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Minimal Residual Disease (*MRD*) for B-ALL	1-2
5	G11S01T06	**Flowcytometry for Minimal Residual Disease (*MRD*) for T-ALL**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Minimal Residual Disease (*MRD*) for T-ALL	1-2
6	G11S01T07	**Flowcytometry for Minimal Residual Disease (*MRD*) for CLL**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Minimal Residual Disease (*MRD*) for CLL	1-2
7	G11S01T08	**Flowcytometry for Minimal Residual Disease (*MRD*) for Lymphoma**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Minimal Residual Disease (*MRD*) for Lymphoma	1-2
8	G11S01T09	**Flowcytometry for Minimal Residual Disease (MRD) for AML**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Minimal Residual Disease (*MRD*) for AML	1-2
9	G11S01T10	**Flowcytometry for Minimal Residual Disease (*MRD*) for MM**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** Flowcytometry for Minimal Residual Disease (*MRD*) for MM	1-2
10	G11S01T11	**Primary Immuno Deficiency (*PID*) Screening panel**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** *T* cells with *CD4* and *CD8* subsets, Naive *T* cell enumeration, Effector and Memory Cells, B cells, *NK* cells and *T* - regulatory cell counts	1-2

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Flowcytometry

BONE MARROW ASPIRATE STUDIES						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
11	G11S01T12	**Lymphocyte subset analysis (*LSSA*) – comprehensive**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** T cells with *CD4* and *CD8* subsets, Naive *T cell* enumeration, Effector and Memory Cells, *B cells*, *NK cells* and *T* - regulatory cell counts	1-2
12	G11S01T13	**PNH by FLAER**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** PNH by FLAER	1-2
13	G11S01T14	**ALPS Screening**	Flowcytometry	3ml BMA/PB in EDTA	**Flowcytometry:** ALPS Screening	1-2

Sarcoma

Lilac Insights™
passion • commitment • genetics

Sarcomas

SARCOMAS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G09S38T01	*EWSR1(22q12)* **translocation**	FISH	FFPE	**FISH:** *EWSR1*(22q12) translocation	7
2	G09S38T02	*EWSR1-FLI1* **translocation**	FISH	FFPE	**FISH:** *EWSR1-FLI1* translocation	7
3	G09S37T01	**Synovial Sarcoma**	FISH	FFPE	**FISH:** t(X;18): *SS18* translocation	7
4	G09S37T02	**Myxoid Sarcoma**	FISH	FFPE	**FISH:** t(12;16): *FUS* translocation	7
5	G09S37T03	**Infantile fibrosarcoma**	FISH	FFPE	**FISH:** *EWSR1(*22q12*)* translocation, FISH: *NTRK3* (15q25*)* translocation	7
6	G09S37T04	**Extra Skeletal Myxoid chondrosarcoma**	FISH	FFPE	**FISH:** *NR4A3(TEC)(*9q22.3*)* translocation	7
7	G09S37T05	**Alveolar Rhabdomyosarcoma**	FISH	FFPE	**FISH:** *FOXO1(*13q14*)* translocation	7
8	G09S37T06	*MDM2* **Gene Amplification**	FISH	FFPE	**FISH:** *MDM2* Gene Amplification	7
9	G10S08T01	*MDM2* **Gene Amplification,** *TP53* **mutation**	FISH + Molecular	FFPE	**FISH:** *MDM2* Gene Amplification, **Molecular:** *TP53* mutation (exon 4-9)	10
10	G09S37T07	*TFE3* **t(Xp11.2) translocation in Alveolar soft tissue sarcoma**	FISH	FFPE	**FISH:** *TFE3 t(*Xp11.2*)* translocation	7
11	G08S19T01	*CTNNB1***(B-Catenin) mutations in Desmoid type Fibrometosis**	Molecular	FFPE	**Molecular:** *CTNNB1*(B-Catenin) mutations hot spots	7-10
12	G08S15T02	*TP53* **mutation sequencing**	Molecular	FFPE	**Molecular:** *TP53* mutation sequencing (exon 4-9)	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Solid Tumors

Lilac Insights™
passion • commitment • genetics

Bladder Cancer and Brain Tumors/Neurological Cancers/Neuroblastoma

BONE MARROW ASPIRATE STUDIES						
S.No.	Lilac Code	Panel Name	Methodology	Specimen Requirements	Tests/Markers	TAT/Working Days
1	G09S36T01	**FISH Panel Bladder Cancer (Urothelial Cancer)**	FISH	Urine in Sterile Container without any additives	**FISH:** Aneuploidy of Chromosome 3, 7, 17 & *del(9p)*	5
2	G09S39T01	**FISH Panel Brain Tumors/Neurological Cancers**	FISH	FFPE Tissue	**FISH:** 11p36 deletion, 19p13 deletion	7
3	G09S40T01	**FISH Panel Neuroblastoma**	FISH	FFPE Tissue	**FISH:** *MYCN* amplification, 1p36 deletion	7

Breast Cancer

BREAST CANCER						
S.No.	Lilac Code	Panel Name	Methodology	Specimen Requirements	Tests/Markers	TAT/Working Days
1	G09S17T01	**FISH for *ERBB2(HER2/neu)* Breast Cancer**	FISH	FFPE Block	**FISH:** *ERBB2 (HER2/neu)* Amplification	7
2	G09S28T03	**Oncoinsights™ FISH *NTRK3 & ETV6* translocation Breast Cancer**	FISH	FFPE Block	**FISH:** *NTRK3* & *ETV6* translocation	7
3	G09S28T02	***NTRK3* translocation**	FISH	FFPE Block	**FISH:** *NTRK3* translocation	7
4	G09S28T01	***ETV6* translocation**	FISH	FFPE Block	**FISH:** *ETV6* translocation	7
5	G08S04T03	**Germline Panel**	NGS	3ml-5ml of Peripheral blood in 2 tubes of EDTA	**NGS:** *BRCA1, BRCA2, TP53* and *ACMG* recommended genes	15-20
6	G08S04T05	***PIK3CA* mutation analysis**	Molecular	3ml PB in EDTA	**Molecular:** *PIK3CA* mutation by Sanger Sequencing	7
7	G08S04T04	**Germline 3 Panel (*HBOC*)**	NGS	3ml-5ml of Peripheral blood in 2 tubes of EDTA	**NGS:** *BRCA1, BRCA2* and *TP53*	15-20
8	G08S04T02	**Germline - *BRCA1 & BRCA2***	NGS	3ml-5ml of Peripheral blood in 2 tubes of EDTA	**NGS:** *BRCA1, BRCA2*	15-20
9	G08S14T02	**56 Gene Test**	NGS	FFPE Block	**NGS:** 56 Genes	15-20
10	G08S14T03	**56 Gene Test - Liquid Biopsy**	NGS	10 ml Peripheral Blood in Streck Tube (2tubes)	**NGS:** 56 Genes	15-20

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Colorectal Cancer

COLORECTAL CANCER						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G08S13T06	**Colorectal Cancer Panel**	Molecular	FFPE Block	**Molecular:** *KRAS, NRAS* by Sanger Sequencing, *BRAF* (ASO-PCR)	6
2	G08S13T12	**Colorectal Cancer Panel with** *PIK3CA*	Molecular	FFPE Block	**Molecular:** *KRAS, NRAS, PIK3CA* mutation by Sanger Sequencing, *BRAF* (ASO-PCR)	6
3	G08S13T13	**Colorectal Cancer Panel with MSI by PCR:** *KRAS, NRAS & BRAF* **Sequencing + MSI by PCR**	Molecular	FFPE Block & PB in EDTA(for PCR	**Molecular:** *KRAS, NRAS* by Sanger Sequencing, *BRAF* (ASO-PCR) & **Molecular:** Microsatellite Instability Index [*MLH-1, MSH-2, MSH-6, PMS-2*] by PCR	6 & 11
4	G08S13T14	**Colorectal Cancer Panel with MSI by** *IHC*: *KRAS, NRAS & BRAF* **Sequencing + MSI by** *IHC*	Molecular IHC	FFPE Block	**Molecular:** *KRAS, NRAS* by Sanger Sequencing, *BRAF* (ASO-PCR) & **Immunohistochemistry:** Microsatellite Instability Index [*MLH-1, MSH-2, MSH-6, PMS-2*] by IHC	6 & 10
5	G08S14T02	**56 Gene Test**	NGS	FFPE Block	**NGS:** 56 Genes	15-20
6	G08S14T03	**56 Gene Test - Liquid Biopsy**	NGS	10 ml Peripheral Blood in Streck Tube (2tubes)	**NGS:** 56 Genes	15-20

INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
7	G08S13T07	**Microsatellite Instability Index by** *IHC*	IHC	FFPE Block	**Immunohistochemistry:** Microsatellite Instability Index by *IHC* [*MLH-1, MSH-2, MSH-6, PMS-2*]	5
8	G08S13T08	**Microsatellite Instability Index by PCR**	Molecular	FFPE Block & PB in EDTA (for PCR)	**Molecular:** Microsatellite Instability Index by PCR [*MLH-1, MSH-2, MSH-6, PMS-2*]	11
9	G08S13T01	*KRAS* **(exon2, exon3) testing**	Molecular	FFPE Block	**Molecular:** *KRAS* (exon2, exon3) testing by Sanger Sequencing	6
10	G08S13T10	*NRAS* **(exon2, exon3) testing**	Molecular	FFPE Block	**Molecular:** *NRAS*(exon2,exon3) testing by Sanger Sequencing	6
11	G08S13T03	*BRAF* **V600E**	Molecular	FFPE Block	**Molecular:** BRAF V600E mutation (*ASO-PCR*)	6

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Colorectal Cancer

INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
12	G08S13T11	*PIK3CA* **mutation analysis**	Molecular	3ml PB in EDTA	**Molecular:** *PIK3CA* mutation by Sanger Sequencing	7
13	G08S13T02	*c-KIT* **(ex 9,11,13,17) mutation sequencing**	Molecular	FFPE Block	**Molecular:** *c-KIT*(ex 9,11,13,17) mutation sequencing	7
14	G08S13T04	*EGFR* **(exon18-21) mutation testing**	Molecular	FFPE Block	**Molecular:** *EGFR* (exon18-21) mutation testing by Sanger sequencing	6
15	G08S13T05	*EGFR* **(exon18-21) mutation & PDL1 by IHC**	Molecular	FFPE Block	**Molecular:** *EGFR* (exon18-21) mutation testing by Sanger sequencing & **Immunohistochemistry:** PDL1 testing by IHC	6
16	G08S13T09	*EGFR* **(exon18-21) mutation &** *KRAS* **(exon2, exon3) testing**	Molecular	FFPE Block	**Molecular:** *EGFR* (exon18-21) mutation & *KRAS* (exon2, exon3) testing	6

Gastrointestinal Stromal Tumors

GASTROINTESTINAL STROMAL TUMORS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G08S18T02	*KIT* **GIST panel**	Molecular	FFPE Block	**Molecular:** *KIT* (exons 9, 11, 13, 17) mutations by Sanger Sequencing	6
2	G08S18T01	*PDGFRA* **Sequencing test**	Molecular	FFPE Block	**Molecular:** *PDGFRA* by Sanger Sequencing	6
3	G08S14T02	**56 Gene Test**	NGS	FFPE Block	**NGS:** 56 Genes	15-20
4	G08S14T03	**56 Gene Test - Liquid Biopsy**	NGS	10 ml Peripheral Blood in Streck Tube (2tubes)	**NGS:** 56 Genes	15-20
5	G08S17T01	**FISH** *ERBB2 (Her2/Neu)* **Gastric Cancer**	FISH	FFPE Block	**FISH:** *ERBB2*(Her2/Neu) amplification in Gastric Cancer	7

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Melanoma

MELANOMA						
S.No.	Lilac Code	Panel Name	Methodology	Specimen Requirements	Tests/Markers	TAT/Working Days
1	G08S08T01	*BRAF* **V600E**	Molecular	FFPE Block	**Molecular:** *BRAF* V600E mutation(*ASO-PCR*)	6
2	G08S14T02	**56 Gene Test**	NGS	FFPE Block	**NGS:** 56 Genes	15-20

Miscellaneous

MISCELLANEOUS						
S.No.	Lilac Code	Panel Name	Methodology	Specimen Requirements	Tests/Markers	TAT/Working Days
1	G08S15T23	*TP53* **Sequencing on** *FFPE*	Molecular	FFPE Block	**Molecular:** *TP53* (exons 4-9) by sanger Sequencing	10
2	G08S15T01	*TP53* **Sequencing on** *BMA/PB*	Molecular	3-5ml of BMA/PB in EDTA tube	**Molecular:** *TP53* (exons 4-9) by sanger Sequencing	10
3	G08S15T02	*TP53* **Sequencing on** *BMA/PB*	Molecular	3-5ml of BMA/PB in EDTA tube	**Molecular:** *TP53* (exons 2-11) by sanger Sequencing	10
4	G08S15T19	**Oncoinsights™Thyroid Cancer Panel**	NGS	FFPE (>20% Tumor Nuclei with a minimum tumor surface area of 5mm * 5mm (25mm2)	**NGS:** *EGFR,BRAF, KRAS, NRAS, HRAS, RET-PTC, PAX8-PPAR* gamma, *TSC1, TSC2*	24
5	G09S37T06	*MDM2* **Amplification in solid tumors**	FISH	FFPE Block	**Molecular:** *MDM2* Amplification	7
6	G10S08T01	*MDM2* **Amplification,** *TP53* **mutation in solid tumors**	Molecular	FFPE Block	**Molecular:** *MDM2* Amplification, *TP53* mutation (exon 4-9)	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Non-small Cell Lung Cancer (NSCLC)

NON-SMALL CELL LUNG CANCER(NSCLC)						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
1	G10S05T01	**FISH Combipanel *NSCLC***	FISH + Molecular	FFPE Block (Preferably Tumor Area Marked)	**FISH:** *ALK* Translocation, *ROS1* translocation **Molecular:** *EGFR* (exon (18-21) mutations by sequencing.	10
2	G09S16T06	**Oncoinsights™ FISH Combipanel-1 NSCLC W/O MET**	FISH + Molecular+IHC	FFPE Block (Preferably Tumor Area Marked)	**FISH:** *ALK* Translocation, *ROS1* translocation, **Molecular:** *EGFR* (exon (18-21) mutations by sequencing, **IHC:** *PDL1*	10
3	G10S05T02	**Oncoinsights™ FISH Combipanel-1 NSCLC**	FISH + Molecular+IHC	FFPE Block (Preferably Tumor Area Marked)	**FISH:** *ALK* Translocation, *ROS1* translocation, MET amplification **Molecular:** *EGFR* (exon (18-21) mutations by sequencing, **IHC:** *PDL1*	10
4	G10S05T03	**Oncoinsights™ FISH Combipanel-2 NSCLC**	FISH + Molecular+IHC	FFPE Block (Preferably Tumor Area Marked)	**FISH:** *ALK* Translocation, *ROS1* translocation, MET amplification **Molecular:** *EGFR* (exon (18-21) mutations, *KRAS* (exon 2,3) mutations by sequencing, *BRAF V600E* mutation (*ASO PCR*), **IHC:** *PDL1*	10
5	G10S05T04	**Non-Small cell lung cancer (*ALK+EGFR*)**	FISH + Molecular	FFPE Block (Preferably Tumor Area Marked)	**FISH:** *ALK* Translocation, **Molecular:** *EGFR* (exon (18-21) mutations by sequencing.	6
6	G08S12T09	**Non-Small cell lung cancer (*EGFR+ KRAS*)**	Molecular	FFPE Block (Preferably Tumor Area Marked)	**Molecular:** *EGFR* (exon (18-21) mutations, *KRAS* (exon2,3) mutation by sequencing.	6
7	G08S14T02	**56 Gene Test**	NGS	FFPE Block	**NGS:** 56 Genes	15-20
8	G08S14T03	**56 Gene Test - Liquid Biopsy**	NGS	10 ml Peripheral Blood in Streck Tube (2 tubes)	**NGS:** 56 Genes	15-20
9	G08S15T06	**NSCLC *NGS* Panel (*FFPE*)**	NGS	FFPE Block (Preferably Tumor Area Marked)	**NGS:** *HER2, PIK3CA, EGFR,BRAF, KRAS,* Translocation: *ALK, ROS1, RET,* Copy Number Variation: *MET,NTRK1,NTRK2, NTRK3*	15
10	G08S15T10	**NSCLC *NGS* Liquid Biopsy**	NGS	10 ml Peripheral Blood in Streck Tube (2tubes)	**NGS:** *ALK, BRAF, EGFR, ERBB2, KRAS, MAP2K1, MET, NRAS, PIK3CA, ROS1,* and *TP53*	15

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

Non-small Cell Lung Cancer (NSCLC)

INDIVIDUAL MARKERS						
S.No.	**Lilac Code**	**Panel Name**	**Methodology**	**Specimen Requirements**	**Tests/Markers**	**TAT/Working Days**
11	G08S12T02	*KRAS* **mutation testing**	Molecular	FFPE Block	**Molecular:** *KRAS* (exon 2,3) mutation testing by Sanger Sequencing	6
12	G08S12T01	*EGFR* **(exon 18-21) mutation testing**	Molecular	FFPE Block	**Molecular:** *EGFR* (exon 18-21) mutation testing by Sanger sequencing	6
13	G10S06T01	*PDL1* **test by** *IHC* **(Roche Ventanna Platform)**	IHC	FFPE Block	**IHC:** *PDL1* (Roche Ventanna FDA Approved Platform)	10
14	G08S15T11	*PDL1* **test by** *IHC* **(DAKO Platform)**	IHC	FFPE Block	**IHC:** *PDL1* (*DAKO* Platform)	10
15	G09S16T02	*ALK* **Translocation**	FISH	FFPE Block	**FISH**: *ALK* Translocation	5
16	G09S16T03	*ROS1* **Translocation**	FISH	FFPE Block	**FISH**: *ROS1* Translocation	5
17	G09S16T05	*MET* **Amplification**	FISH	FFPE Block	**FISH**: *MET* Amplification	5
18	G09S37T06	*MDM2* **Amplification**	FISH	FFPE Block	**FISH**: *MDM2* Amplification	7
19	G08S12T03	*EGFR* **(exon 18-21) mutation & PDL1 by IHC**	Molecular + IHC	FFPE Block	**Molecular:** *EGFR* (exon 18-21) mutation testing by Sanger sequencing & **IHC:** *PDL1*	6
20	G08S12T04	*c-KIT* **(ex 9,11,13,17) mutation sequencing**	Molecular	FFPE Block	**Molecular:** *c-KIT* (ex 9,11,13,17) mutation sequencing	7
21	G08S12T05	**Molecular test** *HCL* **(BRAFV600E mutation)**	Molecular	FFPE Block	**Molecular:** *BRAF V600E* mutation (*ASO-PCR*)	6
22	G08S12T06	**Liquid Biosy -** *EGFR* **sense test**	ddPCR	10 ml Peripheral Blood in Streck Tube (2 tubes)	**ddPCR:** *EGFR*-Sense test (*Del 19 & L858R*)	10
23	G08S12T07	**Liquid Biosy -** *EGFR* **resist test**	ddPCR	10 ml Peripheral Blood in Streck Tube (2 tubes)	**ddPCR:** *EGFR* - Resist test (*T790M*)	10
24	G08S12T08	**Liquid Biosy -** *EGFR* **Lung panel**	ddPCR	10 ml Peripheral Blood in Streck Tube (2 tubes)	**ddPCR:** *EGFR*-Sense test (*Del 19 & L858R*) *EGFR* - Resist test(*T790M*)	10

FISH: FISH Studies Molecular: Molecular Studies IHC: IHC Studies Flowcytometry: Flowcytometry Studies ddPCR: ddPCR Studies NGS: NGS Studies

Collect specimen as per the test information. Note: All FFPE blocks – tumor area should be marked or H&E slides with tumor area marked should be provided.

COLLECTION AND TRANSPORTATION OF SPECIMENS FOR ONCOGENETICS STUDIES

Oncocytogenetics

Hematolymphoid Malignancies:

- Bone Marrow Aspirate (BMA) / Peripheral Blood (PB) is the recommended tissue for FISH. BMA is highly recommended for karyotyping test.
- In absence of BMA,PB is accepted for karyotyping provided PB contains ⩾40% blasts. Collect 3-5ml of bone marrow aspirate/5-8 ml of peripheral blood into a sterile Sodium heparin vacutainer (Green top).
- **In case of drytap bone marrow, we accept minimum 2 unmounted bone marrow smear slides for FISH analysis.**
- In Multiple myeloma FISH studies, 5-6ml of BMA is highly recommended for purification of plasma cells followed by FISH.
- Clinical history should include provisional diagnosis, Flow report, disease status whether at diagnosis, on treatment, primary AML, MDS, secondary/transformed AML, MDS.
- For chromosome breakage studies, 6-10ml of PB in sodium Heparin vacutainer is required for patient as well as for unrelated sex-, age –matched, non-alcoholic, non-smoker, healthy control.

Transportation:
BMA/PB/liquid specimens placed in zip-lock bag along with TRF which should be transported at room temp (RT) or with a cool pack in hot environment and should reach to service lab within 24-48 hrs after collection.

Solid Tumors:
- FFPE tissue block with marked tumor area along with H&E slide
- TRF should include detail clinical history with provisional diagnosis, histopathology, IHC details.

Transportation:
FFPE tissue block/s along with H & E slides are placed in zip-lock bag and transferred to the service laboratory at Room Temperature.
Bladder Cancer FISH: Urine specimen (20-40ml in a sterile plastic container)

Molecular Oncology

Hematolymphoid Malignancies:

Bone Marrow Aspirate (BMA) / Peripheral Blood is the recommended tissue. Collect 3-5ml of bone marrow aspirate/5ml of peripheral blood into a sterile EDTA vacutainer (Purple top). Collect 10 ml peripheral blood in case of monitoring BCR-ABL1 transcript copies when patient is on treatment.

Transportation:
BMA/PB/liquid specimens should be transported with a cool pack and should reach to service lab within 24-48 hrs after collection.

Solid Tumors:
- FFPE tissue block with marked tumor area along with H&E slide
- TRF should include detail clinical history with provisional diagnosis, histopathology, IHC details.

Transportation:
FFPE tissue block/s along with H & E slides are placed in zip-lock bag and transferred to the service laboratory at Room Temperature.

** TRF should be labelled with **biohazard tag** in case of infectious disease history.

Note:
Lilac Insights is continuously validating and introducing new tests, to check the availability of Oncogenetics tests that are not mentioned in the list, kindly contact Dr. Pratibha Kadam Amare, Chief-Cancer and Clinical Genetics (Contact: +91- 9869249494) or Dr. Hrishikesh Lele, Sr. Scientific Officer, Oncocytogenetics Lab (Contact: +91- 7045900435), Molecular Oncology staff (Contact: +91- ,) or Srinivas, Product Manager (+91-7045695164) Or LILAC INSIGHTS +91 22 41841438, 2241841448.
Management of Lilac Insights Private Limited has the right to discontinue or change the panel of any tests mentioned in the list or change prices without prior notification.

All the TAT is from the receipt of the sample in the Lab.

Oncoinsights are the panels created with vast experience of Dr. Pratibha Kadam Amare and latest guidelines (like NCCN, ELN, WHO, ASH & ASCO) updates in cancer genetics for better and accurate risk stratification of the disease.

List of Offerings from Lilac Insights

Cytogenetic Tests	Cytogenetic Tests	Molecular genetics
Hematological Malignancies	*Solid Tumors*	*Hematological Malignancies*
Chronic Myeloid Leukemia - Chronic Phase/Accelerated Phase/ Blast Phase	Breast Cancer	Acute Myeloid Leukemia
Acute Myeloid Leukemia	Gastric Cancer	Chronic Myeloid Leukemia
Acute Promyelocytic Leukemis	Non-Small Cell Lung Cancer	Acute Promyelocytic Leukemia
Myelodysplactic Syndromes	Neuroblastoma	Acute Lymphoblastic Leukemia
Myeloproliferative Neoplasms	Brain Tumors	Myeloproliferative Neoplasms
Acute Lymphoblastic Leukemia(B-Cell/T-Cell)	Neurological Cancers	Myelodysplastic Syndromes
Chronic Lymphocytic Leukemia	Ewing Sarcoma	Early T Cell precursor - Acute Lymphoblastic Leukemia
Sex-mismatched Bone Marrow Transplant	Alveolar Rhabdomyosarcoma	Chronic Lymphocytic Leukemia
Chronic Eosinopholic Leukemia	Extraskeletal Myxoid Chondrosarcoma	Hairy Cell Leukemia
Ph1 Like B- Acute Lymphoblastic Leukemia	Infantile Fibrosarcoma	*Solid Tumors*
Multiple Myeloma on Purified Plasma Cells	Myxoid Liposarcoma	Non Small Cell Lung Cancer
Hairy cell Leukemia	Synovial sarcoma	Colorectal Cancer
Lymphoma FISH Panels	Bladder Cancer	Gastrointestinal Stromal Tumors
IGH Translocations		Melanoma
Mantle Cell Lymphoma		*TP53* Deletion
Follicular Lymphoma		
Burkitt Lymphoma		
Diffuse large B-Cell Lymphoma		
Dual Hit Lymphoma(High Grade B-Cell Lymphoma)		
Triple Hit Lymphoma(High Grade B-Cell Lymphoma)		
MALT Lymphoma		
Lymphoma B-Cell		
Lymphoma T-Cell		
Hepatosplenic γ, δ T-cell Lymphoma		

For Further Details, contact us at:
Dr. P. S. Kadam Amare: +91 9869 249 494
Mr. A B Srinivas - +91 7045 695 164
Tel: +91 22 41841438 | Fax: +91 22 41841448

Why Lilac Insights™

Advances in genetics will create a healthier tomorrow. With a continuous emphasis on innovation and technology, Lilac Insights shall keep contributing to progression of genetics in healthcare and research. We will remain customer-focused. We will keep reinventing our processes, develop skills and create opportunities for genetic professionals. We will not only improve access to advanced genetic solutions in India but also be seen as a collaborator of choice by global organizations, seeking quality genetic services at affordable costs.

Contact us:
LILAC INSIGHTS PVT. LTD
301-302, Rupa Solitaire Premises Co-Op Society Ltd., Sector-1, Building A-1, Millennium Business Park, MIDC, Navi Mumbai - 400710.
Tel: +91 22 41841438 | Fax: +91 22 41841448
Email:info@lilacinsights.com www.lilacinsights.com